



## REVIEW ARTICLES

# Peptides and current methods on bovine tuberculosis diagnosis

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### KEYWORDS

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**Abstract:** Bovine tuberculosis (bTB) is a respiratory disease caused by *Mycobacterium bovis* that mainly infects cattle and adversely affects animal health and the livestock economy. Additionally, bTB affects human health as a zoonotic disease. Therefore, implementing diagnostic tests and slaughter policies campaigns is a valuable strategy to control this disease. The tuberculin skin test (TST) and the interferon-gamma (IFN- $\gamma$ ) assay are applied as current *ante-mortem* bTB diagnostic approaches. In addition, the choice of antigens is critical for the bTB diagnostic technique. The sensitivity of currently TST tests range from 40 to 95%. Thus, available tests present limitations with methodology and/or antigen used. In this scenario, several antigens are derived from inactivated cells or proteins, although peptides have been increasingly studied and used due to their numerous advantages. From this perspective, this review provides an overall literature review of the current *ante-mortem* bTB diagnostic tests, their advantages and limitations, and the peptide antigens used to improve test performance.

### Highlights

- The incidence of Bovine tuberculosis in different countries and regions were cataloged;
- The main aspects of the current ante-mortem diagnostic of bTB are discussed;
- The remain problems of bTB and the need to develop efficient diagnostic tests are highlighted;
- The tests that employ peptide antigens were the focus of the discussion.

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**Table 1.** Incidence of bovine tuberculosis in different regions and countries. The cattle population and disease prevalence data were gathered from the Food and Agriculture Organization of the United Nations (FAO) database, official agriculture and livestock organizations, and studies carried out in specific cities or regions.

Region	Country	Bovine livestock population in 2019 (million numbers)	Prevalence of bTB (%)	Reference
Asia	Bangladesh	24.1	45.6	Islam et al. (2020)
	China	63.5	2.4	Gong et al. (2021)
	India	193.4	7.3	Srinivasan et al. (2018)
Africa	Ethiopia	63.2	51.2	Kemal et al. (2019)
	Morocco	3.3	57.7	Azami et al. (2018)
Europe	Spain	6.6	2.3	MAPA, España (2020)
	England	9.4	5.5	Animal and Plant Health Agency (2020)
	Ireland	6.5	4.2	Department of Agriculture Food and the Marine (2021)
	Italy	6.4	3.3	Abbate et al. (2020)
North America	United States of America	94.8	<0.001	U.S. Department of Agriculture (2017)
	Canada	11.5	1.6	World Organization for Animal Health (2019c)
South America	Brazil	214.6	1.3	MAPA, Brazil (2020)
	Uruguay	11.4	0.02	MGAP, Uruguay (2019)
	Argentina	54.4	0.46	Garcia et al. (2021)
Oceania	Australia	24.7	0.01	Sergeant et al. (2017)
	New Zealand	10.1	0.09	OSPRI New Zealand (2016)

10 million diagnostic tests for bTB were performed in Brazil between 2014 and 2018 MAPA, Brazil (2020). In Uruguay, control strategies aiming to improve bTB control use a combination of two different tests, regular testing every three months and mathematical modeling (Picasso-Risso et al., 2021). In effect, accurate and early diagnosis of bTB is a key factor in determining its prevalence and epidemiological status, enabling authorities to establish preventive measures and eradicate the disease.

Tests currently being used presents limitations of accuracy which can lead to false diagnosis and/or false idea of the epidemiological status. From this perspective, this review discusses the major *ante-mortem* diagnostic techniques for bTB, their current applications and limitations, with a focus on the test antigens, especially synthetic peptides, by providing an overview on merging approaches, test parameters, and platforms.

### Bovine tuberculosis *ante-mortem* diagnosis

Cell-mediated response plays a central role during *M. bovis* infection, and tuberculin and IFN- $\gamma$  assays are used to detect this response (Lyashchenko et al., 2020). On the other hand, humoral marker detection tests such as ELISA can be applied to identify animals in advanced phases of infection and tested negative in the tuberculin test. In this context, humoral marker detection tests can be used to complement the results of cellular response tests (Garbaccio et al., 2019).

A compilation of diagnostic tests for bTB screening is listed (Table 2), and the specifications of each are given in the subsequent sections.

### Tuberculin (TST)

The primary diagnostic test for cattle TB is the Tuberculin (TST). The Purified Protein Derivative (PPD) is used as the antigen for the test and comprises several antigens derived from *M. bovis*. The test is based on the delayed-type hypersensitivity reaction. In order to perform TST, PPD is inoculated into the skin of the neck or caudal fold and the skin thickness is measured before and after inoculation (World Organization for Animal Health, 2018). The result is interpreted as negative if the thickness is  $\leq 2$  mm and the animals show no clinical symptoms, e.g., edema, exudation, inflammation, or pain of the lymphatic ducts at the region of application. Animals with clinical symptoms or skin thickness  $> 4$  mm are interpreted as positive. Tuberculin, which was first developed by Koch at the end of the nineteenth century, is an important tool for diagnosis and eradication programs in many countries. However, PPD comprises a mixture of proteins, lipids, and carbohydrates from an *M. bovis* (World Organization for Animal Health, 2018) AN5 culture. Therefore, PPD specificity markedly decreases once the animals are sensitized to environmental nonpathogenic mycobacteria, which may contain some antigens present in PPD (Roperto et al., 2017). Furthermore, the effectiveness of tuberculin varies between various manufacturers as its composition has not been completely characterized. TST

**Table 2.** Overview of the latest diagnostic tests and methods for bovine tuberculosis: sensitivity, specificity and antigen used. Horizontal lines divide the table in test category, as the same antigen can be presented in different tests. Commercial assays such as ELISA INDEXX and IGRA BOVIGAM can be found with symbols alongside their sensitivity and specificity.

Test	Evaluation type	Antigen	Sensitivity - Specificity (%)	Reference
Tuberculin (TST)	Skin thickness, cellular response	PPD	63.70 - <i>not shown</i>	Casal et al. (2017)
		PPD	87.6- 83.6	Singhla et al. (2019)
		Peptides of ESAT6, CFP10, Rv3615c (DST)	76 - <i>not shown</i>	Srinivasan et al. (2019)
		7 recombinant proteins, Rv3616c peptide cocktail (MDT)	71-100 - 100	Middleton et al. (2021)
IGRA	IFN- $\gamma$ levels	PPD	85- 90.4 <sup>†</sup>	Al-Mouqatea et al. (2018)
		Peptides of ESAT6, CFP10, Rv3615c (DST)	<i>not shown</i>	Srinivasan et al. (2019)
		Peptide cocktail of ESAT6, CFP10	76.2 - 96.5	Picasso-Risso et al. (2019)
		Peptide cocktails: ESAT-6/CFP-10 and Rv3615c	91 - 96	Coad et al. (2019)
		7 recombinant proteins, Rv3616c peptide cocktail (MDT)	100 - 97-100	Middleton et al. (2021)
ELISA	Antibody detection	MBP70, MPB83 recombinant proteins	63- 98 <sup>‡</sup>	Waters et al. (2011)
		Recombinant proteins, MBP70 peptide	93.1 - 98.4%	Whelan et al. (2008)

<sup>†</sup>BOVIGAM commercial test; <sup>‡</sup>INDEXX ELISA commercial test

performance varies depending on different factors, e.g., the dose applied, PPD preparation, the site of application, and data interpretation. It can be observed high variability in values of sensitivity in different studies, such as 63.70% (Casal et al., 2017), 75.3-95.2% with a median of 87.6% (Singhla et al., 2019) and a variation between 40.1% and 92.2% in the same study (Casal et al., 2012). A variability on specificity is sometimes presented as well, between 74.2% and 92.8% (Singhla et al., 2019). Hence, ongoing efforts are being conducted to identify new antigens not present in environmental mycobacteria by selecting proteins that belong to the MTC, especially to *M. bovis*. Moreover, TST should be associated with low costs to allow its widespread application.

An optimal diagnostic test should differentiate infected from vaccinated animals (DIVA). Srinivasan et al., (2019) developed a novel TST (DST) with antigens derived from specific *M. bovis* proteins. The test consists of 13 peptides representing these three antigens: ESAT-6, CFP10, and ESX-1 (Rv3615c) and can identify a higher percentage of naturally infected animals than the PPD test. DST could identify DIVA after vaccination (Srinivasan et al., 2020). Synthetic peptides that mimic the Rv3616c antigen have been included with other protein antigens to develop a new TST test for bovine tuberculosis, called MDT (Middleton et al., 2021). The test showed 100% of specificity and greater sensitivity than the DST previously mentioned, with relative sensitivities of 100% and 73% for the MDT and DST, respectively in experimentally infected cattle, and 71% and 29% respectively in naturally-infected cattle. Moreover, a comparison with different numbers of residues peptides was made (20 versus 40-mer), and the MDT of 40-mer peptides induced a strong skin test response (Middleton et al., 2021).

### Interferon-Gamma Release Assay (IGRA)

The IGRA or IFN- $\gamma$  test is an alternative diagnostic test for bTB. IGRA has been used in parallel with TST in many countries to prevent tuberculosis outbreaks (Guétin-Poirier et al., 2020; Jang et al., 2020). In order to perform IGRA, the blood from sampled animals is incubated with antigens to stimulate the production of IFN- $\gamma$  by sensitized T lymphocytes (World Organization for Animal Health, 2018). Then, the IFN- $\gamma$  levels are measured using a sandwich enzyme immunoassay. The BOVIGAM<sup>®</sup> (Prionics AG, Schlieren, Switzerland) IFN- $\gamma$  assay is a commercial test that contains antigens from bovine tuberculin PPD. Field studies have reported that IFN- $\gamma$  is more sensitive than TST, with similar or lower specificity (Keshavarz et al., 2016). Studies performed between 1991 and 2006 revealed that the sensitivity and specificity values of BOVIGAM<sup>®</sup> were 87.6 and 96.6%, respectively, based on 15 field studies (de la Rua-Domenech et al., 2006). On the other hand, recent studies have reported that the sensitivity and specificity of the BOVIGAM<sup>®</sup> test ranged between 78-85% and 90.4-91.4% (Al-Mouqatea et al., 2018; Picasso-Risso et al., 2019), respectively.

The BOVIGAM<sup>™</sup> PC-EC stimulating antigen (Prionics AG, Schlieren-Zurich, Switzerland) is a commercial IGRA based on a peptide cocktail derived from ESAT-6 and CFP10. Compared to the PPD test, the commercial test showed slightly lower sensitivity and higher specificity (Thermo Fisher Scientific, 2019). Furthermore, in a study comparing the accuracy of IGRA tests, the one with peptide antigens derived from ESAT-6/CFP10 demonstrated higher sensitivity and specificity (Picasso-Risso et al., 2019). Srinivasan et al. (2019) tested DST (peptide-based TST using ESAT-6, CFP10, and Rv3615c proteins). The cocktail of synthetic peptides showed enhanced performance compared to a construction containing these



three antigens as a fusion protein in IGRA and skin tests. Similarly, the previously mentioned MDT was evaluated for its potential to stimulate IFN- $\gamma$  production (Middleton et al., 2021). The MDT test, composed of recombinant proteins and a cocktail of synthetic peptides, induced higher levels of IFN- $\gamma$  than the DST test in experimentally and naturally infected animals (100% sensitivity in both groups). In addition, non-infected control animals showed a strong response when tested with avian and bovine PPD and no response for MDT, showing great specificity.

Studies have focused on adding other biomarkers alongside IFN- $\gamma$  to better detect *M. bovis*. Two peptide cocktails, ESAT-6/CFP-10 and Rv3615c, were tested in comparison with avian and bovine PPD to measure the IFN- $\gamma$  and CXCL10 readouts (Coad et al., 2019). Although the CXCL10 measuring sensitivity was lower than IFN- $\gamma$  (which was 100%) with any antigens, the new biomarker could complement sensitivity by identifying TST-positive animals that did not have an IFN- $\gamma$  readout when using peptide cocktails as antigens.

### Enzyme-linked Immuno Sorbent Assay (ELISA)

ELISA is widely used for diagnosing many human and animal diseases. The test is based on detecting antibodies against *M. bovis*, which prevail in advanced stages of the infection, thus providing an indirect way to diagnose bTB. Different antigens and methods have been investigated for ELISA-based bTB diagnosis since the 1980s, and antigens such as MBP70, ESAT-6, CFP10, and MBP83 have been examined to establish a reliable ELISA for bTB (Sun et al., 2021). However, developing antibody assays entails various limitations, including decreased sensitivity, especially in samples from early infection stages. Several studies have attempted to increase the performance of ELISA. The ELISA IDEXX *M. bovis* Ab Test (IDEXX, Maine, USA), a commercial kit for field testing, detects antibodies against MPB70 and MPB83 proteins and can be used as a supplementary test in bTB eradication programs (Paulo Alex Machado Carneiro et al., 2021). The sensitivity and specificity of the test have shown values of 63 and 98%, respectively. However, sensitivity can be increased with TST (Waters et al., 2011).

A synthetic peptide-based ELISA can be developed by peptide-based capture, wherein peptides are directly immobilized in wells of the microtiter plate by the adsorption procedure (Pandey et al., 2021), which was the case of a study that synthesized peptides derived from the known immunogenic MBP70 protein. However, peptides were tested alongside other recombinant proteins, and the ELISA test results showed sensitivity and specificity values of 93.1% and 98.4%, respectively (Whelan et al., 2008). Another study explored peptides from different regions of MBP70, recognized by anti-MBP70 chicken antibodies, but no further tests were conducted with cattle sera (Modise, 2012).

### Peptide antigens

The use of synthetic peptides for diagnostic purposes has increased in the past few years since they are highly pure and can be chemically modified (Pandey et al., 2021).

Many immunological and bioinformatic techniques exist to identify, select and design immunodominant peptides. They are used to screen the pathogen proteins and search within them for the most antigenic regions. In this way, it is possible to reduce the time needed to develop a diagnostic test and to find the antigens that will result in tests with the best performance. Some online tools available for linear B-cell epitope prediction are ABCPred, Bepipred 2.0, among others. Ellipro and Discotope are available for prediction of discontinuous or conformational B-cell epitopes (both available through The Immune Epitope Database - IEDB). Immunological approaches are based on the interaction between antibody and antigen, e.g., PEPSCAN (Lorenzo et al., 2021), phage display (Ramli et al., 2019) and combinatorial peptide libraries (Bozovičar & Bratkovič, 2019). Meanwhile, as mentioned, the bioinformatic tools make use of amino acids properties and epitopes databases to predict immunogenic regions (Bai et al., 2018; Ortega-Tirado et al., 2020). Hence, validating new peptide markers for diagnostic purposes has become more accessible, less expensive, and more effective.

Peptides can be easily synthesized using the standard solid-phase method, in which molecules are covalently bonded to solid support material, and amino acids are synthesized step-by-step using Fmoc protective groups. Highly pure amino acids with Fmoc are available for this purpose (>99%). The constant improvement in protection strategies during synthesis and increasing purity levels have made peptides more accessible in terms of industrial-scale use (Behrendt et al., 2016). Other approaches are being explored in peptide production, e.g., micro-flow technology, which has a shorter production time, fewer risks with dangerous compound reagents, and ready scale-up with high reproducibility (Fuse et al., 2018). Using synthetic peptides as antigens is cost-effective since their manufacture overcomes the limitations associated with using recombinant proteins, expressed in heterologous systems (*i.e.*, *E. coli*) such as formation of inclusion bodies (Bhatwa et al., 2021) with its laborious steps (purification and refolding). These synthetic molecules can be evaluated individually or as a cocktail of peptides with one or more antigens, simplifying experimental validation. Furthermore, whole proteins can be replaced with peptides to reduce nonspecific binding, thus evaluating the immune response for a single antigenic epitope (Vedova-Costa et al., 2021), improving test sensitivity, a major problem in bTB assays as cross-reactions with the environment and MTC bacteria are common.

In the last few years, several studies have highlighted the potential of synthetic peptides in diagnostic tests for various diseases, e.g., bovine anaplasmosis (Quiroz-Castañeda et al., 2019), bovine coxiellosis (Yadav et al., 2020), leishmaniosis (Link et al., 2017), capro-ovine listeriosis (Malla et al., 2021), toxoplasmosis (Alves et al., 2019), sensorineural hearing loss caused by cytomegalovirus (Zavaglio et al., 2021), and COVID-19 (Polvere et al., 2022). The accuracy values for most of these test's range between 60 and 100% for sensitivity and 73 and 100% for specificity.

### Overview

This review discusses the methods currently used for diagnosing of bTB, their sensitivity/specificity, and

advantages/disadvantages. Bovine tuberculosis is a major worldwide problem. In summary, the research articles discussed here presented studies of different uses of the characterized proteins such as ESAT-6 and/or CFP10, like novel approaches with peptides of these antigens. In addition, some articles explored the use of Rv3615c in the peptide form, but rarely the MBP70 and/or MBP83 proteins. Commonly, it is used defined groups for evaluating the test accuracy, which are generally cattle natural infected by *M. bovis* and noninfected cattle control from tuberculosis-free farms. The number of animals used for the standardization of tests ranged from 25 to 279. For the TST, the antigen is inoculated in experimentally infected cattle to evaluate its efficacy. Samples taken are different for each test, as sera is used for ELISA and Peripheral Blood Mononuclear Cells (PBMCs) are used for IGRA. Sensitivity and specificity were calculated using commercially available statistics program such as Prism 4 and 7. Within these programs different statistical approaches were used: analysis of variance repeated measures, Tukey's multiple comparisons test, Student t test and Bayesian approach; all of those with the 95% confidence interval (CI).

It can be observed that although PPD delivers great specificity, it stills lack sensitivity in most tests. TST using PPD as an antigen provides variable results as it can be seen in different articles presenting values with wide differences. Also, it presents the problem of cattle being sensitized by environmental mycobacteria, which may be resolved with the use of more specific antigens. Therefore, the TST is rapid and easy-managing, but lacks novel antigens to increase its performance. In the last few years, ELISA has been the most studied method by many research groups since the current commercial tests lack a suitable performance. Nevertheless, the antigens used are recombinant proteins, and if peptides are included, they are usually the already known ESAT-6/CFP10 cocktails. Although not sufficiently explored, other peptide antigens could present satisfactory results and constitute a good alternative. In this scenario, immunoinformatics can help identify and select new peptide candidates, whereas peptide-manufacturing technologies are widespread, making them more accessible. In addition, it is now easier and less expensive to efficiently test novel antigens. Hence, it is necessary to research these antigens and develop simple, rapid, and sensitive diagnostic methods for bTB to mitigate the disease's adverse effects. This is especially true in non-developed regions, where fewer alternatives are available, and new findings could facilitate applying the guidelines for bTB control.

## Conflict of interests

None.

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## References

- Abbate, J. M., Arfuso, F., Iaria, C., Arestia, G., & Lanteri, G. (2020). Prevalence of bovine tuberculosis in slaughtered cattle in Sicily, Southern Italy. *Animals*, *10*(9), 1-11. <http://dx.doi.org/10.3390/ani10091473>. PMID:32839384.
- Al-Mouqatea, S., Alkhamis, M., Akbar, B., Ali, A., Al-Aqeel, H., Bin-Heji, A., Razzaque, M., Alvarez, J., & Perez, A. (2018). Bayesian estimation of ELISA and gamma interferon test accuracy for the detection of bovine tuberculosis in caudal fold test-negative dairy cattle in Kuwait. *Journal of Veterinary Diagnostic Investigation*, *30*(3), 468-470. <http://dx.doi.org/10.1177/1040638718759574>. PMID:29431048.
- Alves, L. M., Barros, H. L. S., Flauzino, J. M. R., Guedes, P. H. G., Pereira, J. M., Fujiwara, R. T., Mineo, T. W. P., Mineo, J. R., de Oliveira, R. J., Madurro, J. M., & Brito-Madurro, G. (2019). A novel peptide-based sensor platform for detection of anti-*Toxoplasma gondii* immunoglobulins. *Journal of Pharmaceutical and Biomedical Analysis*, *175*, 112778. <http://dx.doi.org/10.1016/j.jpba.2019.112778>. PMID:31352171.
- Animal and Plant Health Agency. (2020). *Bovine tuberculosis in England in 2019: Epidemiological analysis of the 2019 data and historical trends*. [https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment\\_data/file/923195/tb-epidemiology-england-2019.pdf](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/923195/tb-epidemiology-england-2019.pdf)
- Azami, H. Y., Ducrottoy, M. J., Bouslikhane, M., Hattendorf, J., Thrusfield, M., Conde-Álvarez, R., Moriyón, I., Zúñiga-Ripa, A., Muñoz Álvaro, P. M., Mick, V., Bryssinckx, W., Welburn, S. C., & Zinsstag, J. (2018). The prevalence of brucellosis and bovine tuberculosis in ruminants in Sidi Kacem Province, Morocco. *PLoS One*, *13*(9), e0203360. <http://dx.doi.org/10.1371/journal.pone.0203360>. PMID:30226847.
- Bai, X., Wang, D., Liu, Y., Xiao, L., Liang, Y., Yang, Y., Zhang, J., Lin, M., & Wu, X. (2018). Novel epitopes identified from *Mycobacterium tuberculosis* antigen Rv2629 induces cytotoxic T lymphocyte response. *Immunology Letters*, *203*, 21-28. <http://dx.doi.org/10.1016/j.imlet.2018.06.005>. PMID:29908955.
- Behrendt, R., White, P., & Offer, J. (2016). Advances in Fmoc solid-phase peptide synthesis. *Journal of Peptide Science*, *22*(1), 4-27. <http://dx.doi.org/10.1002/psc.2836>. PMID:26785684.
- Bhatwa, A., Wang, W., Hassan, Y. I., Abraham, N., Li, X.-Z., & Zhou, T. (2021). Challenges associated with the formation of recombinant protein inclusion bodies in *Escherichia coli* and strategies to address them for industrial applications. *Frontiers in Bioengineering and Biotechnology*, *9*, 630551. <http://dx.doi.org/10.3389/fbioe.2021.630551>. PMID:33644021.
- Bozovičar, K., & Bratkovič, T. (2019). Evolving a peptide: Library platforms and diversification strategies. *International Journal of Molecular Sciences*, *21*(1), 215. <http://dx.doi.org/10.3390/ijms21010215>. PMID:31892275.
- Carneiro, P. A. M., Moura, S., Viana, R. B., Monteiro, B. M., Socorro Lima Kzam, A., de Souza, D. C., Coelho, A. S., Ribeiro Filho, J. D., Jordão, R. S., Tavares, M. R. M., & Kaneene, J. B. (2021). Study on supplemental test to improve the detection of bovine tuberculosis in individual animals and herds. *BMC Veterinary Research*, *17*(1), 137. <http://dx.doi.org/10.1186/s12917-021-02839-4>. PMID:33789652.
- Carneiro, P. A. M., & Kaneene, J. B. (2018). Bovine tuberculosis control and eradication in Brazil: Lessons to learn from the US and Australia. *Food Control*, *93*, 61-69. <http://dx.doi.org/10.1016/j.foodcont.2018.05.021>.
- Casal, C., Bezos, J., Díez-Guerrier, A., Álvarez, J., Romero, B., de Juan, L., Rodríguez-Campos, S., Vordermeier, M., Whelan, A., Hewinson, R. G., Mateos, A., Domínguez, L., & Aranaz, A. (2012). Evaluation of two cocktails containing ESAT-6, CFP-10 and Rv-3615c in the intradermal test and the interferon- $\gamma$  assay for diagnosis of bovine tuberculosis. *Preventive Veterinary Medicine*, *105*(1-2), 149-154. <http://dx.doi.org/10.1016/j.prevetmed.2012.02.007>. PMID:22391021.

- Casal, C., Infantes, J. A., Risalde, M. A., Díez-Guerrier, A., Domínguez, M., Moreno, I., Romero, B., de Juan, L., Sáez, J. L., Juste, R., Gortázar, C., Domínguez, L., & Bezos, J. (2017). Antibody detection tests improve the sensitivity of tuberculosis diagnosis in cattle. *Research in Veterinary Science*, *112*, 214-221. <http://dx.doi.org/10.1016/j.rvsc.2017.05.012>. PMID:28521256.
- Coad, M., Doyle, M., Steinbach, S., Gormley, E., Vordermeier, M., & Jones, G. (2019). Simultaneous measurement of antigen-induced CXCL10 and IFN- $\gamma$  enhances test sensitivity for bovine TB detection in cattle. *Veterinary Microbiology*, *230*, 1-6. <http://dx.doi.org/10.1016/j.vetmic.2019.01.007>. PMID:30827373.
- de la Rúa-Domenech, R., Goodchild, A. T., Vordermeier, H. M., Hewinson, R. G., Christiansen, K. H., & Clifton-Hadley, R. S. (2006). Ante mortem diagnosis of tuberculosis in cattle: A review of the tuberculin tests,  $\gamma$ -interferon assay and other ancillary diagnostic techniques. *Research in Veterinary Science*, *81*(2), 190-210. <http://dx.doi.org/10.1016/j.rvsc.2005.11.005>. PMID:16513150.
- Department of Agriculture Food and the Marine, Ireland (2021). *National bovine TB statistics*. <https://www.gov.ie/en/publication/5986c-national-bovine-tb-statistics-2020/>
- Djafar, Z. R., Benazi, N., Bounab, S., Sayhi, M., Diouani, M. F., & Benia, F. (2020). Distribution of seroprevalence and risk factors for bovine tuberculosis in east Algeria. *Preventive Veterinary Medicine*, *183*, 105127. <http://dx.doi.org/10.1016/j.prevetmed.2020.105127>. PMID:32905887.
- Fuse, S., Otake, Y., & Nakamura, H. (2018). Peptide synthesis utilizing micro-flow technology. *Chemistry, an Asian Journal*, *13*(24), 3818-3832. <http://dx.doi.org/10.1002/asia.201801488>. PMID:30341812.
- Garbaccio, S. G., Garro, C. J., Delgado, F., Tejada, G. A., Eirin, M. E., Huertas, P. S., Leon, E. A., & Zumárraga, M. J. (2019). Enzyme-linked immunosorbent assay as complement of intradermal skin test for the detection of *Mycobacterium bovis* infection in cattle. *Tuberculosis*, *117*, 56-61. <http://dx.doi.org/10.1016/j.tube.2019.05.006>. PMID:31378269.
- García, M. S., Melo, A. F., Carvalho, G. F., Pomim, G. P., Neves, P. M. de S., Silva, R. A. B., de Oliveira, R. O., & Frias, D. F. R. (2021). Epidemiology of bovine tuberculosis in South America. *Research Social Development*, *10*(9), e8610917936. <http://dx.doi.org/10.33448/rsd-v10i9.17936>.
- Gong, Q.-L., Chen, Y., Tian, T., Wen, X., Li, D., Song, Y.-H., Wang, Q., Du, R., & Zhang, X.-X. (2021). Prevalence of bovine tuberculosis in dairy cattle in China during 2010-2019: A systematic review and meta-analysis. *PLoS Neglected Tropical Diseases*, *15*(6), e0009502. <http://dx.doi.org/10.1371/journal.pntd.0009502>. PMID:34138867.
- Guétin-Poirier, V., Rivière, J., Crozet, G., & Dufour, B. (2020). Assessment of the cost-effectiveness of alternative bovine tuberculosis surveillance protocols in French cattle farms using the mixed interferon gamma test. *Research in Veterinary Science*, *132*, 546-562. <http://dx.doi.org/10.1016/j.rvsc.2020.08.005>. PMID:32829191.
- Islam, S. K. S., Rumi, T. B., Kabir, S. M. L., van der Zanden, A. G. M., Kapur, V., Rahman, A. K. M. A., Ward, M. P., Bakker, D., Ross, A. G., & Rahim, Z. (2020). Bovine tuberculosis prevalence and risk factors in selected districts of Bangladesh. *PLoS One*, *15*(11), e0241717. <http://dx.doi.org/10.1371/journal.pone.0241717>. PMID:33170869.
- Jang, Y. H., Kim, T. W., Jeong, M. K., Seo, Y. J., Ryoo, S., Park, C. H., Kang, S., Lee, Y. J., Yoon, S. S., & Kim, J. M. (2020). Introduction and application of the interferon- $\gamma$  assay in the national bovine tuberculosis control program in South Korea. *Frontiers in Veterinary Science*, *7*, 222. <http://dx.doi.org/10.3389/fvets.2020.00222>. PMID:32411741.
- Kemal, J., Sibhat, B., Abraham, A., Terefe, Y., Tulu, K. T., Welay, K., & Getahun, N. (2019). Bovine tuberculosis in eastern Ethiopia: Prevalence, risk factors and its public health importance. *BMC Infectious Diseases*, *19*(1), 39. <http://dx.doi.org/10.1186/s12879-018-3628-1>. PMID:30630431.
- Keshavarz, R., Mosavari, N., Geravand, M. M., Tadayon, K., Pajoohi, R. A., Solemani, K., Ameri, M., & Gilani, G. V. (2016). Interferon- $\gamma$  assay, a high-sensitivity, specific and appropriate method for detection of bovine tuberculosis in cattle. *International Journal of Mycobacteriology*, *5*(Suppl 1), S219. <http://dx.doi.org/10.1016/j.ijmyco.2016.10.014>. PMID:28043566.
- Kuria, J. K. N. (2019). Diseases caused by bacteria in cattle: Tuberculosis. In H. Kaoud (Ed.), *Bacterial cattle diseases*. London: IntechOpen. <https://doi.org/10.5772/INTECHOPEN.82051>.
- Link, J. S., Alban, S. M., Soccol, C. R., Pereira, G. V. M., & Thomaz Soccol, V. (2017). Synthetic peptides as potential antigens for cutaneous leishmaniasis diagnosis. *Journal of Immunology Research*, *2017*, 5871043. <http://dx.doi.org/10.1155/2017/5871043>. PMID:28367456.
- Lorenzo, M. A., Pachón, D., Maier, A., Bermúdez, H., Losada, S., Toledo, M., Pujol, F. H., Alarcón de Noya, B., Noya, O., & Serrano, M. L. (2021). Immunoinformatics and pepscan strategies on the path of a peptide-based serological diagnosis of COVID19. *Journal of Immunological Methods*, *495*, 113071. <http://dx.doi.org/10.1016/j.jim.2021.113071>. PMID:33991531.
- Lyashchenko, K. P., Vordermeier, H. M., & Waters, W. R. (2020). Memory B cells and tuberculosis. *Veterinary Immunology and Immunopathology*, *221*, 110016. <http://dx.doi.org/10.1016/j.vetimm.2020.110016>. PMID:32050091.
- Malla, B. A., Ramanjeneya, S., Vergis, J., Malik, S. S., Barbudde, S. B., & Rawool, D. B. (2021). Comparison of recombinant and synthetic listeriolysin- O peptide- based indirect ELISA vis-à-vis cultural isolation for detection of listeriosis in caprine and ovine species. *Journal of Microbiological Methods*, *188*, 106278. <http://dx.doi.org/10.1016/j.mimet.2021.106278>. PMID:34246691.
- MAPA, Brazil, Ministério da Agricultura Pecuária e Abastecimento. (2020). *Diagnóstico situacional do Programa de Controle e Erradicação da Brucelose e da Tuberculose Animal*. <https://www.gov.br/agricultura/pt-br/assuntos/sanidade-animal-e-vegetal/saude-animal/programas-de-saude-animal/pncebt/DSPNCEBT.pdf>
- Mapa, España, Ministerio de Agricultura, Pesca y Alimentación. (2020). *Programa Nacional de Erradicación de Tuberculosis Bovina presentado por España para el año 2020*. [https://www.mapa.gob.es/es/ganaderia/temas/sanidad-animal-higiene-ganadera/pnetb\\_2020final\\_tcm30-523317.PDF](https://www.mapa.gob.es/es/ganaderia/temas/sanidad-animal-higiene-ganadera/pnetb_2020final_tcm30-523317.PDF)
- Middleton, S., Steinbach, S., Coad, M., McGill, K., Brady, C., Duignan, A., Wiseman, J., Gormley, E., Jones, G. J., & Vordermeier, H. M. (2021). A molecularly defined skin test reagent for the diagnosis of bovine tuberculosis compatible with vaccination against Johne's Disease. *Scientific Reports*, *11*(1), 2929. <http://dx.doi.org/10.1038/s41598-021-82434-7>. PMID:33536465.
- Modise, B. M. (2012). *Mycobacterium tuberculosis complex-specific antigens for use in serodiagnosis of bovine tuberculosis* [Thesis]. University of Pretoria.
- Ortega-Tirado, D., Niño-Padilla, E. I., Arvizu-Flores, A. A., Velazquez, C., Espitia, C., Serrano, C. J., Enciso-Moreno, J. A., Sumoza-Toledo, A., & Garibay-Escobar, A. (2020). Identification of immunogenic T-cell peptides of *Mycobacterium tuberculosis* PE\_PGRS33 protein. *Molecular Immunology*, *125*, 123-130. <http://dx.doi.org/10.1016/j.molimm.2020.06.026>. PMID:32659597.
- OSPRI New Zealand. (2016). *Annual review 2015/2016*. <https://www.ospri.co.nz/assets/Documents/20201210-TBfree-NOP-2020.pdf>
- Pandey, S., Malviya, G., & Chottova Dvorakova, M. (2021). Role of peptides in diagnostics. *International Journal of Molecular Sciences*, *22*(16), 8828. <http://dx.doi.org/10.3390/ijms22168828>. PMID:34445532.
- Pérez-Morote, R., Pontones-Rosa, C., Gortázar-Schmidt, C., & Muñoz-Cardona, Á. I. (2020). Quantifying the economic impact of bovine tuberculosis on livestock farms in South-Western Spain. *Animals*, *10*(12), 2433. <http://dx.doi.org/10.3390/ani10122433>. PMID:33353111.
- Picasso-Risso, C., Alvarez, J., VanderWaal, K., Kinsley, A., Gil, A., Wells, S. J., & Perez, A. (2021). Modelling the effect of test-and-slaughter strategies to control bovine tuberculosis in endemic



- high prevalence herds. *Transboundary and Emerging Diseases*, 68(3), 1205-1215. <http://dx.doi.org/10.1111/tbed.13774>. PMID:32767833.
- Picasso-Risso, C., Perez, A., Gil, A., Nunez, A., Salaberry, X., Suanes, A., & Alvarez, J. (2019). Modeling the accuracy of two in-vitro bovine tuberculosis tests using a bayesian approach. *Frontiers in Veterinary Science*, 6, 261. <http://dx.doi.org/10.3389/fvets.2019.00261>. PMID:31457019.
- Polvere, I., Voccola, S., Cardinale, G., Fumi, M., Aquila, F., Parrella, A., Madera, J. R., Stilo, R., Vito, P., & Zotti, T. (2022). A peptide-based assay discriminates individual antibody response to SARS-CoV-2. *Genes & Diseases*, 9(1), 275-281. <http://dx.doi.org/10.1016/j.gendis.2021.01.008>. PMID:33564711.
- Quiroz-Castañeda, R. E., Tapia-Uriza, T. R., Valencia Mujica, C., Rodríguez-Camarillo, S. D., la Preciado-De Torre, J. F., Amaro-Estrada, I., & Cobaxin-Cárdenas, M. (2019). Synthetic peptides-based indirect ELISA for the diagnosis of bovine anaplasmosis. *International Journal of Applied Research in Veterinary Medicine*, 17(2), 65-70.
- Ramli, S. R., Moreira, G. M. S. G., Zantow, J., Goris, M. G. A., Nguyen, V. K., Novoselova, N., Pessler, F., & Hust, M. (2019). Discovery of *Leptospira* spp. seroreactive peptides using ORFome phage display. *PLoS Neglected Tropical Diseases*, 13(1), e0007131. <http://dx.doi.org/10.1371/journal.pntd.0007131>. PMID:30677033.
- Roperto, S., Varano, M., Russo, V., Lucà, R., Cagiola, M., Gaspari, M., Ceccarelli, D. M., Cuda, G., & Roperto, F. (2017). Proteomic analysis of protein purified derivative of *Mycobacterium bovis*. *Journal of Translational Medicine*, 15(1), 68. <http://dx.doi.org/10.1186/s12967-017-1172-1>. PMID:28372590.
- Sergeant, E., Happold, J., & Langstaff, I. (2017). Evaluation of australian surveillance for freedom from bovine tuberculosis. *Australian Veterinary Journal*, 95(12), 474-479. <http://dx.doi.org/10.1111/avj.12648>. PMID:29243239.
- Singhla, T., Boonyayatra, S., Chulakasian, S., Lukkana, M., Alvarez, J., Sreevatsan, S., & Wells, S. J. (2019). Determination of the sensitivity and specificity of bovine tuberculosis screening tests in dairy herds in Thailand using a Bayesian approach. *BMC Veterinary Research*, 15(1), 149. <http://dx.doi.org/10.1186/s12917-019-1905-x>. PMID:31096976.
- Srinivasan, S., Easterling, L., Rimal, B., Niu, X. M., Conlan, A. J. K., Dudas, P., & Kapur, V. (2018). Prevalence of bovine tuberculosis in India: A systematic review and meta-analysis. *Transboundary and Emerging Diseases*, 65(6), 1627-1640. <http://dx.doi.org/10.1111/tbed.12915>. PMID:29885021.
- Srinivasan, S., Jones, G., Veerasami, M., Steinbach, S., Holder, T., Zewude, A., Fromsa, A., Ameni, G., Easterling, L., Bakker, D., Juleff, N., Gifford, G., Hewinson, R. G., Martin Vordermeier, H., & Kapur, V. (2019). A defined antigen skin test for the diagnosis of bovine tuberculosis. *Science Advances*, 5(7), eaax4899. <http://dx.doi.org/10.1126/sciadv.aax4899>. PMID:31328169.
- Srinivasan, S., Subramanian, S., Shankar Balakrishnan, S., Ramaiyan Selvaraju, K., Manomohan, V., Selladurai, S., Jothivelu, M., Kandasamy, S., Gopal, D. R., Kathaperumal, K., Conlan, A. J. K., Veerasami, M., Bakker, D., Vordermeier, M., & Kapur, V. (2020). A defined antigen skin test that enables implementation of bcg vaccination for control of bovine tuberculosis: Proof of concept. *Frontiers in Veterinary Science*, 7, 391. <http://dx.doi.org/10.3389/fvets.2020.00391>. PMID:32793643.
- Sun, L., Chen, Y., Yi, P., Yang, L., Yan, Y., Zhang, K., Zeng, Q., & Guo, A. (2021). Serological detection of *Mycobacterium tuberculosis* complex infection in multiple hosts by one universal ELISA. *PLoS One*, 16(10), e0257920. <http://dx.doi.org/10.1371/journal.pone.0257920>. PMID:34618810.
- Thermo Fisher Scientific. (2019). *BOVIGAM™ PC-EC Stimulating Antigen (lyophilized)*. <https://www.thermofisher.com/order/catalog/product/7600100#/7600100>
- Trost, B., Stuber, T., Surujballi, O., Nelson, J., Robbe-Austerman, S., Smith, N. H., Desautels, L., Tikoo, S. K., & Griebel, P. (2016). Investigation of the cause of geographic disparities in IDEXX ELISA sensitivity in serum samples from *Mycobacterium bovis*-infected cattle. *Scientific Reports*, 6(1), 22763. <http://dx.doi.org/10.1038/srep22763>. PMID:26949166.
- U.S. Department of Agriculture. (2017). *National Tuberculosis Eradication Program*. <https://www.ers.usda.gov/topics/animal-products/cattle-beef/statistics-information/>
- Uruguay, Ministerio de Ganadería Agricultura y Pesca. (2019). Últimos datos de Tuberculosis Bovina. [http://www.smvu.com.uy/noticias\\_355-tuberculosis-bovina.html](http://www.smvu.com.uy/noticias_355-tuberculosis-bovina.html)
- Vedova-Costa, J. M. D., Ramos, E. L. P., Boschero, R. A., Ferreira, G. N., Soccol, V. T., Santiani, M. H., Pacce, V. D., Lustosa, B. P. R., Vicente, V. A., & Soccol, C. R. (2021). A review on COVID-19 diagnosis tests approved for use in Brazil and the impact on pandemic control. *Brazilian Archives of Biology and Technology*, 64(spe), 1-14. <http://dx.doi.org/10.1590/1678-4324-75years-2021200147>.
- Waters, W., Buddle, B., Vordermeier, H., Gormley, E., Palmer, M., Thacker, T., Bannantine, J., Stabel, J., Linscott, R., Martel, E., Milian, F., Foshaug, W., & Lawrence, J. (2011). Development and evaluation of an enzyme-linked immunosorbent assay for use in the detection of bovine tuberculosis in cattle. *Clinical and Vaccine Immunology; CVI*, 18(11), 1882-1888. <http://dx.doi.org/10.1128/CVI.05343-11>. PMID:21918115.
- Whelan, C., Shuralev, E., O'Keefe, G., Hyland, P., Kwok, H. F., Snoddy, P., O'Brien, A., Connolly, M., Quinn, P., Groll, M., Watterson, T., Call, S., Kenny, K., Duignan, A., Hamilton, M. J., Buddle, B. M., Johnston, J. A., Davis, W. C., Olwill, S. A., & Clarke, J. (2008). Multiplex immunoassay for serological diagnosis of *Mycobacterium bovis* infection in cattle. *Clinical and Vaccine Immunology; CVI*, 15(12), 1834-1838. <http://dx.doi.org/10.1128/CVI.00238-08>. PMID:18927068.
- World Organization for Animal Health - OIE. (2018). *Bovine tuberculosis: Manual of diagnostic tests and vaccines for terrestrial animals*. <https://www.oie.int/app/uploads/2021/03/3-04-06-bovine-tb.pdf>
- World Organization for Animal Health - OIE. (2019a). Panorama 2019-1: Bovine tuberculosis: Global distribution and implementation status of prevention and control measures according to WAHIS data. *Bulletin de l'OIE*, 2019(1), 9-11. <http://dx.doi.org/10.20506/bull.2019.1.2912>
- World Organization for Animal Health - OIE. (2019b). *Roadmap to zoonotic tuberculosis*. [https://www.oie.int/fileadmin/Home/eng/Our\\_scientific\\_expertise/docs/pdf/Tuberculosis/Roadmap\\_zoonotic\\_TB.pdf](https://www.oie.int/fileadmin/Home/eng/Our_scientific_expertise/docs/pdf/Tuberculosis/Roadmap_zoonotic_TB.pdf)
- World Organization for Animal Health - OIE. (2019c). Panorama 2019-1: Bovine tuberculosis in Western Canada (2016): Case response overview. *Bulletin de l'OIE*, 2019(1), 1-3. <http://dx.doi.org/10.20506/bull.2019.1.2928>.
- Yadav, J. P., Malik, S. V. S., Dhaka, P., Kumar, M., Sirsant, B., Gourkhede, D., Barbudde, S. B., & Rawool, D. B. (2020). Comparison of two new in-house Latex Agglutination Tests (LATs), based on the DnaK and Com1 synthetic peptides of *Coxiella burnetii*, with a commercial indirect-ELISA, for sero-screening of coxiellosis in bovines. *Journal of Microbiological Methods*, 170, 105859. <http://dx.doi.org/10.1016/j.mimet.2020.105859>. PMID:32027926.
- Zavaglio, F., Fiorina, L., Suárez, N. M., Fornara, C., De Cicco, M., Cirasola, D., Davison, A. J., Gerna, G., & Lilleri, D. (2021). Detection of genotype-specific antibody responses to glycoproteins b and h in primary and non-primary human cytomegalovirus infections by peptide-based ELISA. *Viruses*, 13(3), 399. <http://dx.doi.org/10.3390/v13030399>. PMID:33802390.